Instructions for handling irradiated *Drosophila* feeder cells

*Updated: April 2021*

Irradiated feeder cells (at least 200 million cells) arrive in a T-75 flask containing approximately 250 mL of Shield’s and Sang M3 Insect Medium supplemented with Bactopeptone, Yeast Extract and 10% Fetal Calf Serum (no antibiotics).

Due to travel and handling, these semi-adherent cells are likely to be suspended in the media when they arrive.

Upon arrival, please handle the cells under aseptic conditions as follows:

1. Remove the cell suspension from the flask, transfer to 50 mL conical tubes.
2. Centrifuge at 1000 g for 8 minutes to collect cell pellets.
3. Resuspend cells in the culture media, such that the final cell density is between 4-6 million cells/ mL.
4. Replate the cells onto 100 mm culture dishes (5 to 6 culture dishes)
5. Place cells in 25C incubator until ready to use
6. We recommend using the cells for your application the soonest possible.

We use feeder cells to support single cell cloning at a density of 1.5 million cells/ mL.